# organic compounds

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# *a*-Onocerin chloroform hemisolvate

### Roland Fröhlich<sup>a</sup>\* and Guido F. Pauli<sup>b</sup>

<sup>a</sup>Organisch-Chemisches Institut, Westfälische Wilhelms-Universität, Corrensstraße 40, D-48149 Münster, Germany, and <sup>b</sup>Institut für Pharmazeutische Biologie und Phytochemie, Westfälische Wilhelms-Universität, Hittorfstraße 56, D-48149 Münster, Germany

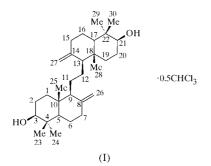
Correspondence e-mail: frohlic@nwz.uni-muenster.de

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The triterpenoid natural product  $\alpha$ -onocerin [8,14-secogammacera-8(26),14(27)-diene-3,21-diol], determined here as the chloroform hemisolvate, C<sub>30</sub>H<sub>50</sub>O<sub>2</sub>·0.5CHCl<sub>3</sub>, consists of two independent symmetric *trans*-decalin C<sub>15</sub> building blocks. Hydrogen bonds between the hydroxyl groups form an infinite two-dimensional network perpendicular to the *c* axis.

### Comment

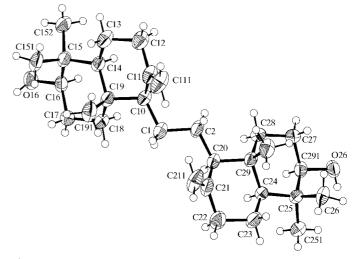
The roots of *Ononis spinosa* L. (Fabaceae) are used medicinally as a diuretic drug. Phytochemical identification, diagnosis of drug adulterations and the standardization of extracts are performed using pure  $\alpha$ -onocerin as a reference compound. While isolated in 1855, its constitution was assigned using chemical methods only in 1955 (Barton & Overton, 1955, and references therein). However, only the structure of the related 8(26),14(27)-diketones have been studied by X-ray analysis so far (Tsuda *et al.*, 1983). We report here the first X-ray structure of a 3,21-onoceradiene-3,21-diol, (I). This is in continuation of the comprehensive spectral evaluation of  $\alpha$ -onocerin focusing on NMR and MS spectral methods which was published recently (Pauli, 2000) and which supported the presence of two *trans*-decalin C<sub>15</sub> building blocks symmetrically linked by a bimethylene bridge.

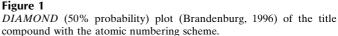


The structural investigation of (I) confirms the assumed geometry. The framework consists mainly of  $Csp^3-Csp^3$  bonds [1.498 (6)–1.576 (5) Å], with only C11–C111 and

C21–C211 [1.317 (7) and 1.329 (7) Å] being clearly  $Csp^2$ –  $Csp^2$  bonds. Also, both bonds to the hydroxyl groups are in the expected range for a single C–O bond [1.445 (5) and 1.441 (5) Å]. Furthermore, the geometries of both *trans*decalin C<sub>15</sub> building blocks are quite similar and comparable with the same skeleton found in gummozine (Nasirov *et al.*, 1977) and 3- $\alpha$ -hydroxymanool hydrate (Kagawa *et al.*, 1993).

Hydrogen bonds between the hydroxyl groups form an infinite two-dimensional network perpendicular to the c axis (Table 1). Besides these, there are no further contacts closer than van der Waals distances.





The recently published NMR spectroscopic evidence for a symmetric constitution of  $\alpha$ -onocerin is now definitively corroborated by the crystallographic data. Therefore, it becomes clear that the C<sub>30</sub> skeleton consists of two halves that are stereochemically identical. Compared with typical pentacyclic triterpene skeletons, such as oleanolic acid, however, this must be rated a very unusual feature with respect to the designated triterpenoid origin of  $\alpha$ -onocerin which has not been supported by biogenetic studies so far. It must be mentioned that no diastereomeric analogues, e.g. isomers with inversion of one or more stereocentres, could be detected by NMR (Pauli, 2000). The biogenetic assignment may be challenged in three points: the necessity for fully stereospecific alterations such as (i) the hydrogenation of the 12,13 double bond and (ii) the hydroxylation of the C-21 position, and most importantly, (iii) an inversion of the decalin ring fusion being [D/E]cis in oleanolic acid. Therefore, the classification of  $\alpha$ -onocerin as a dimeric sesquiterpene is equally justified at this point and should not be omitted. Concerning the numbering of the  $\alpha$ -onocerin framework, which in the literature is again based on the assumption that it represents a triterpene, at this point, revision should be avoided unless the biosynthesis has been investigated.

## **Experimental**

The title compound was obtained from the dried roots of Ononis spinosa (sample No. 9200280, PhytoLab, Vestenbergsgreuth) upon Soxhlet extraction with petrol ether, cleaning-up with active charcoal, in vacuo precipitation, and repeated crystallization from petrol ether, CH<sub>2</sub>Cl<sub>2</sub>/EtOAc (1:1), and CHCl<sub>3</sub> in a final yield of 0.05%. Colourless crystals were obtained from CHCl<sub>3</sub>. Calculated mass for C<sub>30</sub>H<sub>50</sub>O<sub>2</sub>: 442.7. Mass spectrum (El; m/z): 442 (M<sup>+</sup>), 427 ([M-CH<sub>3</sub>]<sup>+</sup>), 409  $([M-CH_3-H_2O]^+)$ , 391  $([M-CH_3-2H_2O]^+)$ , 381  $([M-CH_3-H_2O]^+)$ CO-H<sub>2</sub>O]<sup>+</sup>). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$ , 1.572 (qddd, 1[19]A = eq), 0.952 (ddd, 1[19]B = ax), 1.552 (dddd, 2[20]A = eq), 1.441 (dddd, 2[20]B =ax), 3.090 (dd, 3[21] $\alpha$ ), 0.946 (dd, 5[17]ax), 1.604 ([d]dddd, 6[16]A = eq), 1.239 (dddd, 6[16]B = ax), 2.268 (ddd, 7[15]A = eq), 1.839 (dddddt, 7[15]B = ax), 1.369 (m, 9[13]), 1.369 (m[dddd], 11[12]A),1.055 (m[dddd], 11[12]B), 0.845 (s, 23[30]), 0.616 (s, 24[29]), 0.505 (br s/d, 25[28]), 4.695 (dd, 26[27]A), 4.442 (ddd, 26[27]B). <sup>13</sup>C NMR (CDCl<sub>3</sub>): 36.84 (1[19]), 27.27 (2[20]), 78.43 (3[21]), 38.84 (4[22]), 54.50 (5[17]), 23.81 (6[16]), 38.03 (7[15]), 148.28 (8[14]), 57.28 (9[13]), 39.00 (10[18]), 22.30 (11[12]), 27.91 (23[30]), 15.09 (24[29]), 14.21 (25[28]),106.35 (26[27]).

Crystal data

$C_{30}H_{50}O_2 \cdot 0.5CHCl_3$ $M_r = 502.38$ Orthorhombic, $P2_12_12_1$ a = 7.470 (1) Å b = 14.681 (2) Å c = 27.079 (4) Å V = 2969.7 (7) Å <sup>3</sup> Z = 4 $D_r = 1.124$ Mg m <sup>-3</sup>	Cu K $\alpha$ radiation Cell parameters from 25 reflections $\theta = 22.46-46.14^{\circ}$ $\mu = 1.716 \text{ mm}^{-1}$ T = 223 (2) K Block, colourless $0.30 \times 0.20 \times 0.15 \text{ mm}$
$D_x = 1.124 \text{ Mg m}^{-3}$ Data collection	
Enraf–Nonius CAD-4 diffract- ometer	$\theta_{\max} = 74.29^{\circ}$ $h = 0 \rightarrow 9$

ometer
$\omega/2\theta$ scans
Absorption correction: empirical
<i>via</i> $\psi$ scan (Fair, 1990)
$T_{\min} = 0.627, \ T_{\max} = 0.783$
3450 measured reflections
3450 independent reflections
2318 reflections with $I > 2\sigma(I)$

#### Table 1

Hydrogen-bonding geometry (Å, °).

$D-\mathrm{H}\cdots A$	D-H	$H \cdots A$	$D \cdots A$	$D - H \cdots A$
$O16-H16A\cdots O26^{i}$	0.83	2.04	2.822 (5)	157
O26−H26A···O16 <sup>ii</sup>	0.83	2.17	2.817 (5)	135

 $\begin{array}{l} k = 0 \rightarrow 18 \\ l = 0 \rightarrow 33 \end{array}$ 

3 standard reflections every 250 reflections

frequency: 120 min

intensity decay: 6.7%

Symmetry codes: (i) x, y - 1, z; (ii)  $\frac{1}{2} + x, \frac{3}{2} - y, -z$ .

#### Refinement

<i>,</i>	
Refinement on $F^2$	$w = 1/[\sigma^2(F_o^2) + (0.1687P)^2]$
$R[F^2 > 2\sigma(F^2)] = 0.076$	where $P = (F_o^2 + 2F_c^2)/3$
$wR(F^2) = 0.228$	$(\Delta/\sigma)_{\rm max} = 0.001$
S = 0.991	$\Delta \rho_{\rm max} = 0.70 \ {\rm e} \ {\rm \AA}^{-3}$
3450 reflections	$\Delta \rho_{\rm min} = -0.34 \text{ e } \text{\AA}^{-3}$
313 parameters	Absolute structure: Flack (1983)
H-atom parameters constrained	Flack parameter = $-0.05$ (19)

The asymmetric unit contains half a disordered molecule of chloroform as a residual solvent. This disorder decreases the quality of the whole investigation. The Flack parameter of -0.05 (19) favours the reported absolute configuration, but due to the disorder problem and the fact that a large fraction of Friedel-related reflections have not been measured, this detail should not be overrated. The occupancy of the disordered chloroform was first refined and then fixed with a site-occupancy factor of 0.5. Refinement with anisotropic displacement parameters and/or restraints did not improve the model; therefore, these four atoms are only refined isotropically.

Data collection: *EXPRESS* (Nonius, 1994); cell refinement: *EXPRESS*; data reduction: *MolEN* (Fair, 1990); program(s) used to solve structure: *SHELXS*97 (Sheldrick, 1990); program(s) used to refine structure: *SHELXL*97 (Sheldrick, 1997); molecular graphics: *DIAMOND* (Brandenburg, 1996); software used to prepare material for publication: *SHELXL*97.

Supplementary data for this paper are available from the IUCr electronic archives (Reference: SK1405). Services for accessing these data are described at the back of the journal.

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